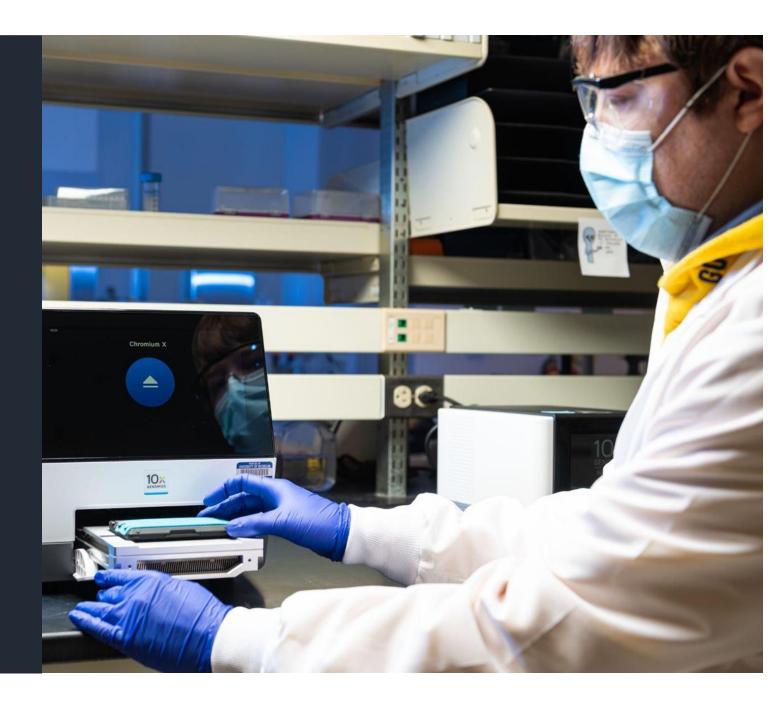
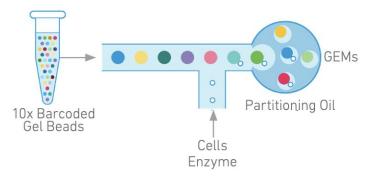
Intro to Single Cell Technologies at the AGC

advanced-genomics@umich.edu

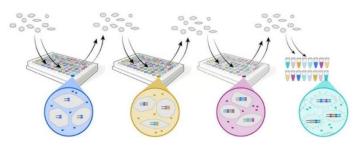


Approaches to Single Cell Sequencing

Microfluidic Droplet-Based



Split-pooling



Benefits:

- throughput
- performance consistency
- low per cell cost
- multi-modal compatible

Limitations:

- size limitation (<30um)
- 3' or 5' bias or targeted



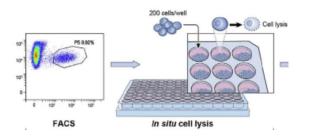
Benefits:

- throughput
- reduced 3' bias
- size agnostic

Limitations:

- fixation
- capture efficiency
- labor intensive

Direct Cell Lysis

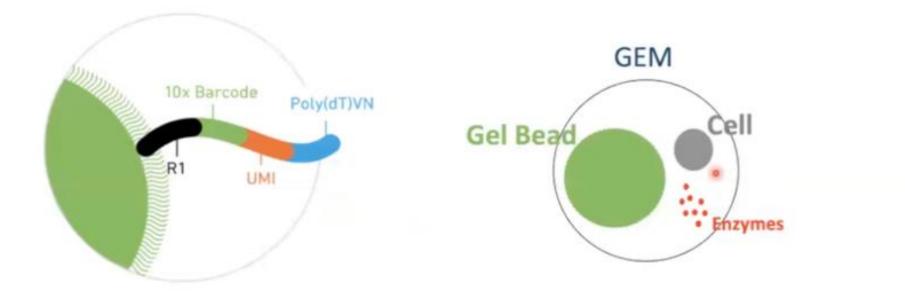


Benefits: - full-length

Limitations:

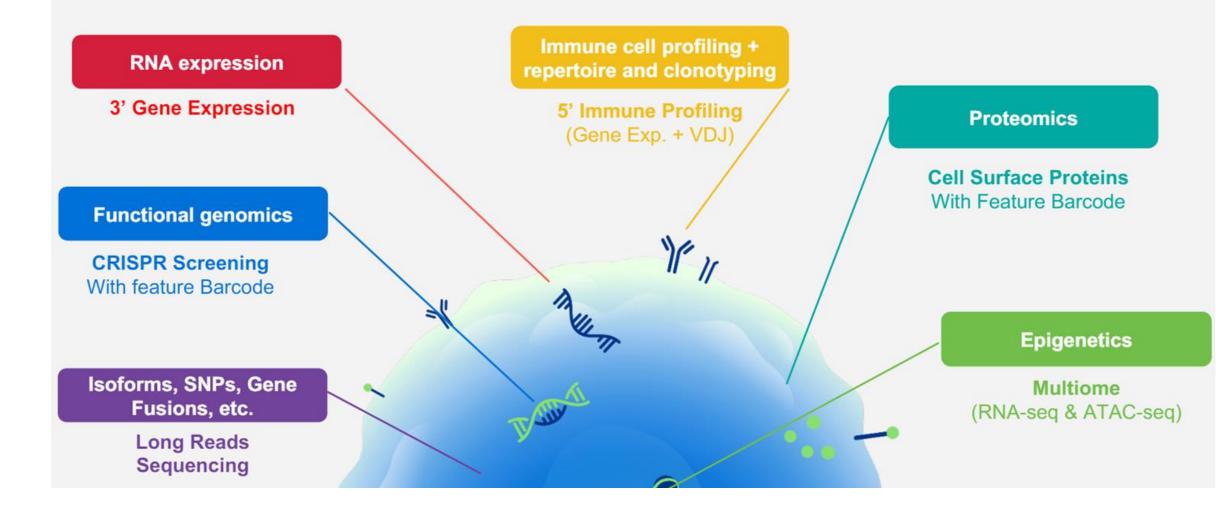
- expense
- throughput

Single Cell RNA Sequencing

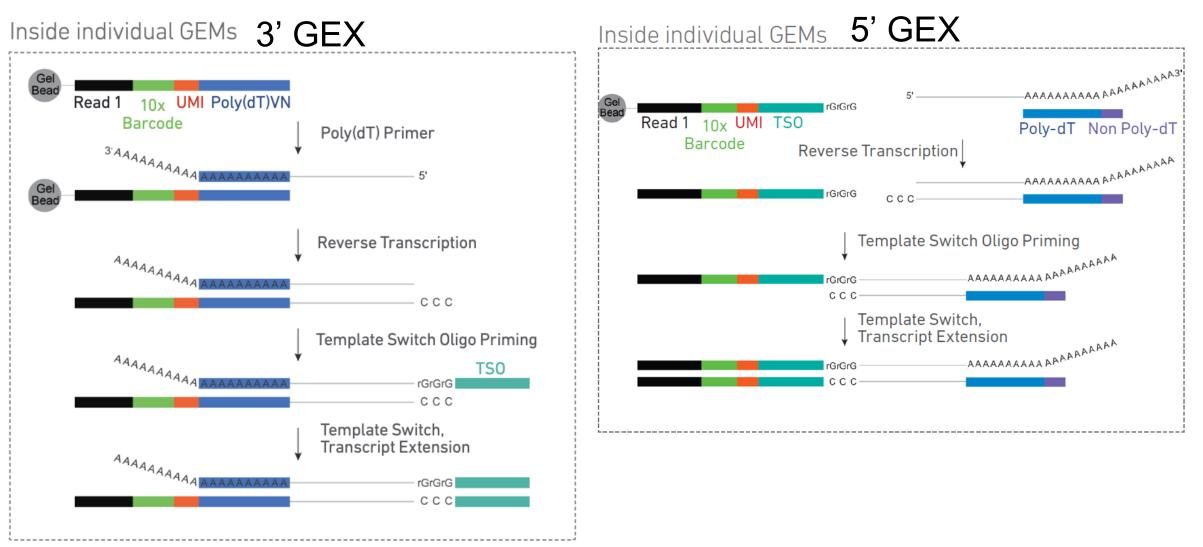




Flavors of Single Cell Sequencing



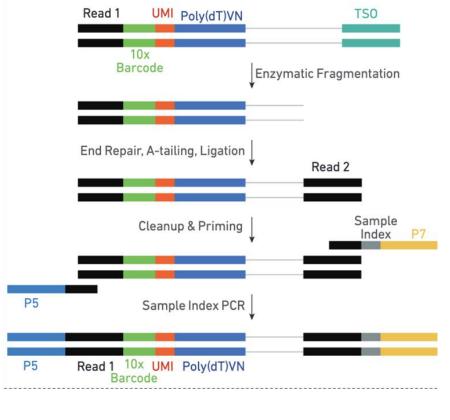
Transcript Capture Mechanisms





10x Capture Mechanisms

Full



Length cDNA **Enzymatic Fragmentation** End Repair, A-tailing, Ligation Read 2 Cleanup & Priming Sample Index i7 Sample Index PCR Sample Sample Index i5 Index i7 P5 Read 1 10x UMI TSO Insert Read 2 BC

Read 1 BC UMI TSO

5' GEX Library

P7

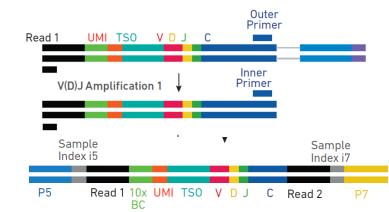
P7

3' GEX Library

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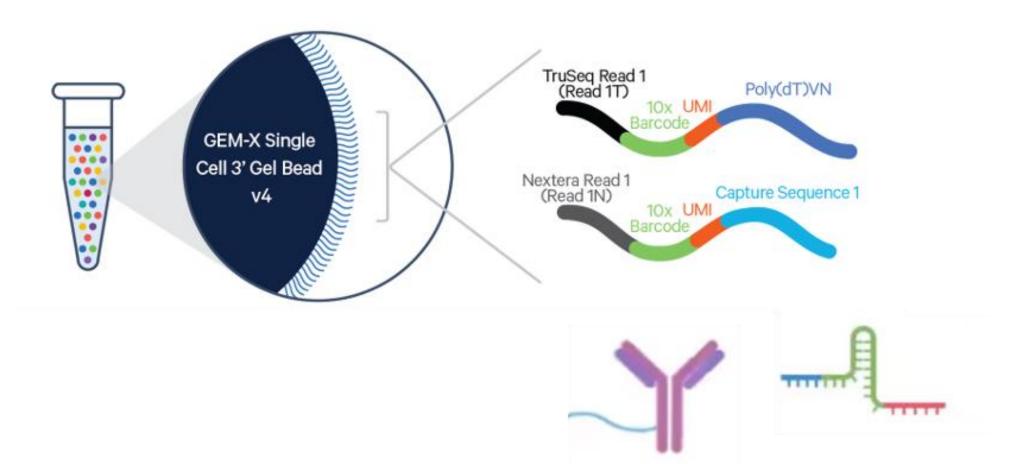
BIOMEDICAL RESEARCH CORE FACILITIES

UNIVERSITY OF MICHIGAN



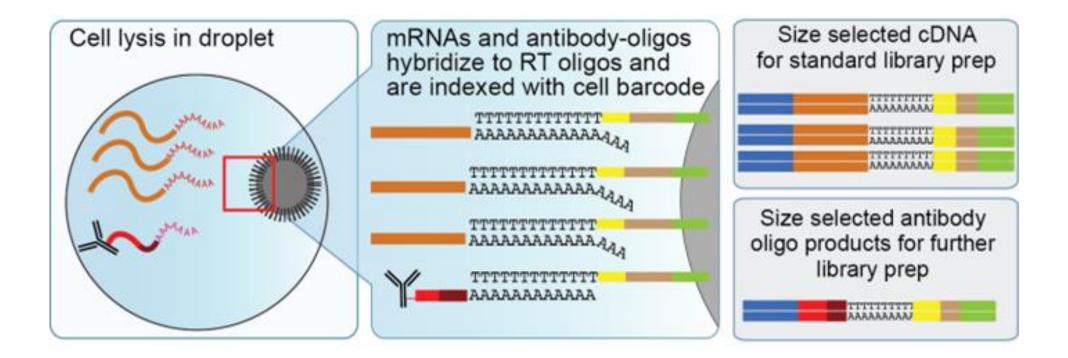
V(D)J Library

Proteomics



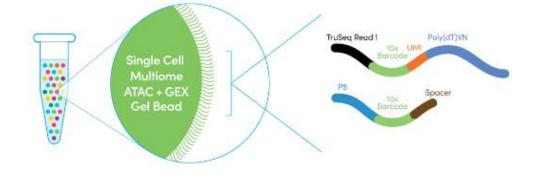


Cellular Indexing of Transcriptomes and Epitopes

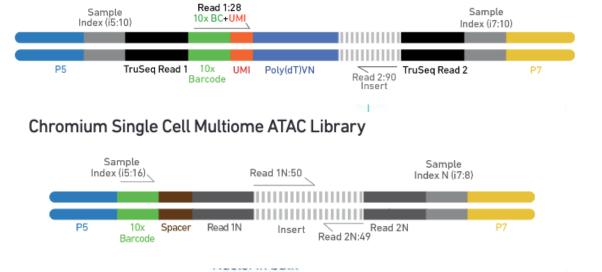




Epi Multiome (ATAC + Gene Expression)



Chromium Single Cell Multiome Gene Expression Library





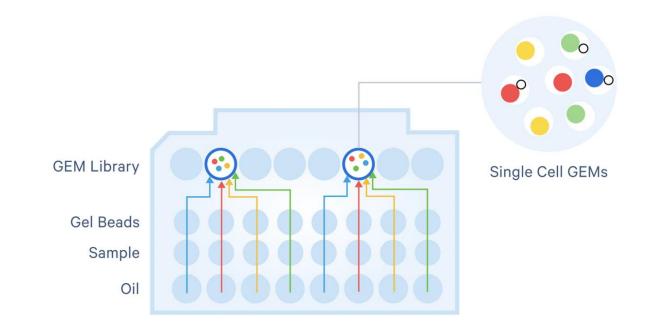
3' and 5' OCM: On-Chip Multiplexing

· Pool 4 samples

- targeting 5000 cells/sample

. Compatible with

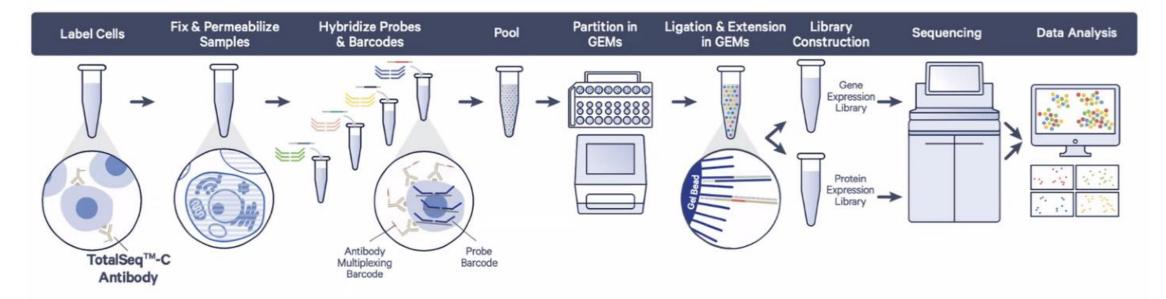
- protein detection
- TCR/BCR profiling
- CRISPR screens





Single Cell RNA Flex System

Same workflow - More cost effective and scalable



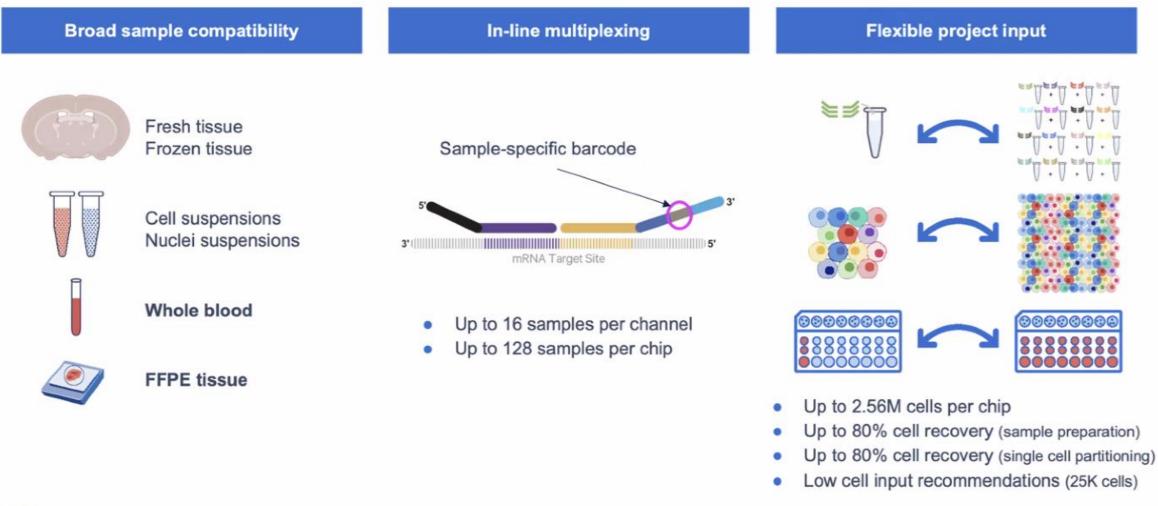
· Sensitive, probe-based whole transcriptome assay

GENOMICS

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- Compatible with Feature Barcode technology for profiling cell surface and intracellular proteins using singleplex or multiplex workflows
- Does not depend on polyA capture; covers more than 18,000 human or mouse genes

GEM-X Flex Gene Expression – any sample at any scale



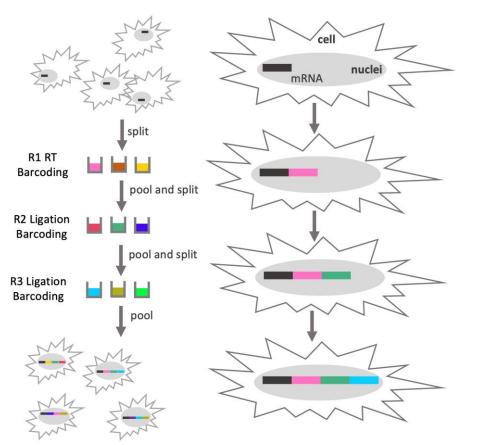


11

SPLiT-seq



- Fix 100K-4 million cells or nuclei
- 1 million cells/96 or
 384 samples
- RNA with TCR and CRISPR add-ons





- Fix 100K-2.5million cells or nuclei
- 500K cells/96 samples
- RNA, Methylation,

ATAC, CRISPR,

Proteomics



Inputs

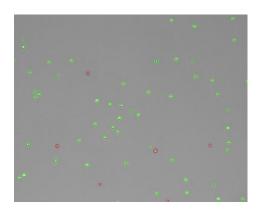
- 10x 3'/5' samples
 - We load 2x as many cells as you target
 - Ideally 1000-1500 cells/ul
 - There is a methanol-mediated fixation protocol _
 - Most common medias are acceptable just keep the cells alive!
 - 10% or less FBS
 - 2% or less BSA
 - NO EDTA
- Flex samples
 - 50mg of frozen or fixed/frozen tissue
 - 10X fixation kit
 - 300k cells/500k nuclei into the fixation —
 - 50k cells/nuclei into hybridization _
 - Two 50um scrolls of FFPE tissue
 - You may need more depending on tissue size/cell density
- Epi Multiome or ATAC

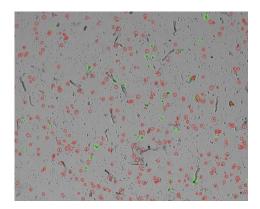
BIOMEDICAL RESEARCH CORE FACILITIES

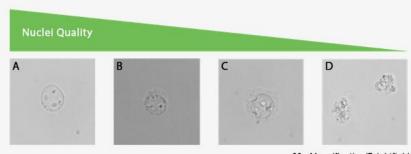
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- Debris-free nuclei suspension
 10x Genomics Buffer (Aliquots available at the AGC)
 Minimum of 3200 nuclei/ul if targeting 10K nuclei for capture
- Split Pooling (Parse/Scale)
 - Fixed cells or nuclei following manufacturer's protocol

GENOMICS CORE







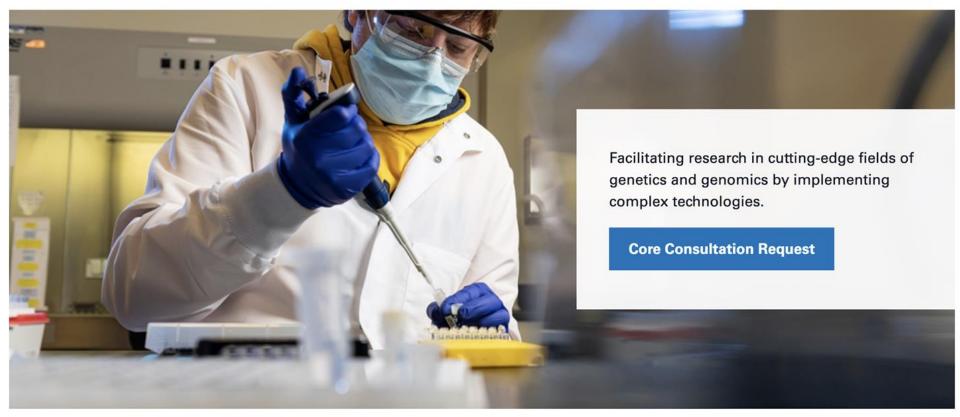
60x Magnification/Brightfield

Single Cell Notes

- FACs sorted cells
 - Usually overestimated by 30-60%
 - Sorting first by ruby and then viability greatly reduces error
- Small population of cells?
 - Concentrate into 30ul of buffer
 - Include an unwanted/abundant population as well
- Run a pilot
 - Start with 1-2 samples to check the data/quality
- Proteome does not equal transcriptome
 - You may sort on a protein but not see the transcript
- Cell target numbers are "in a perfect world"
 - Cell size, tissue/cell type, disease state all affect recovery
 - It may take some optimization to get what you need
- Some samples will fail
 - There will be dropouts whether it is the cell recovery, unexpectedly dead, etc.
 - This is especially true of FFPE samples



ADVANCED GENOMICS CORE



Website: http://michmed.org/agc Email: advanced-genomics@umich.edu

